gradually became yellow. Stirring was continued for 30 min, then the Celite-chloromercuri-2-ethylthio-6-methylauninoparine mixture was filtered, washed with ethanol, and dried *in vacuo* (yield 39.28 g, 99%). The product was further dried and treated with 43.5 mmoles of **3** as described for **1a**. The condensation product was deblocked with methanolic NH₃ as described for **1a**, and **1d** (6.2 g) crystallized during the chloroform extraction of the final aqueous solution. Two recrystallizations from ethanol gave white plates (4.4 g, 30%), mp 206–207.5°.

from ethanol gave white plates (4.4 g, 30%), mp 206-207.5°. *Anal.* Calcd for $C_{13}H_{19}N_5O_4S$: C, 45.73; H, 5.61; N, 20.51. Found: C, 45.79; H, 5.81; N, 20.00.

2-Trifluoromethyl- N° -methyladenosine (1e).--Chloromercuri-2-trifluoromethyl-6-methylaminopurine on Celite was prepared from 4.12 g (19 mmoles) of the purine, 10 g of Celite, 5.18 g (19 mmoles) of HgCl₂, and 0.765 g (19 mmoles) of NaOH as described for 2d; 17.2 g of the chloromercuri-2-trifluoromethyl-6-methylaminopurine-Celite mixture was obtained and treated with 19 mmoles of 3 (from 9.65 g, 19 mmoles, of 5). The procedure and work-up were as described for 1d, and 1e separated from water as a light brown crystalline solid (2.85 g, 43%), mp 203-205°, which crystallized from water as white needles (2.4 g, 36%), mp 203.5-206°. A second recrystallization gave pure 1e, mp 204-206°.

Anal. Calcd for $C_{i2}H_{14}F_3N_5O_4(0.511_2O)$; C, 40.19; H, 4.19; N, 19.54. Found: C, 40.15; H, 4.20; N, 19.49.

2-Chloro- N° **-dimethyladenosine** (1f).—A chloromercuri-2chloro-6-dimethylaminopurine–Celite mixture was prepared in 84% yield, as described for 2d from 6.0 g (30.4 mmoles) of 2-chloro-6-dimethylaminopurine,¹⁷ Celite (12.0 g), HgCl₂(8.25 g, 30.4 mmoles), and NaOH (1.22 g, 30.4 mmoles). The dried product (21.6 g) was treated with 30.4 mmoles of 3 (from 15.3 g, 30.4 mmoles, of 5) as described for 1d. Evaporation of methanolic NH₃ after the deblocking step left a dark oil Trituration with CHCl₃-water gave crystals (4.55 g, 46%) which were recrystallized twice from aqueous ethanol to give 1f as needles (3.5 g), mp 214–215°. Two recrystallizations from methanol gave analytically pure 1f, mp 218–220°.

Anal. Caled for $C_{12}H_{10}ClN_5O_4$: C, 43.60; H, 4.89; N, 21.24. Found: C, 44.02; H, 5.08; N, 21.27.

2-Methoxy- N° -methyladenosine (1c).—Sodium methoxide (2 N, 8 ml) was added to a solution of 1a (1.0 g) in dry methanol (100 ml), and the solution was heated under reflux for 50 hr. Paper chromatography showed that only a trace of 1a remained. The solution was cooled, neutralized with dilute HCl, and evaporated to a white residue. This was extracted with dry ethanol (50 ml), NaCl was filtered off, and the filtrate was evaporated leaving a colorless gum. This was dissolved in water (50 ml), and the solution was filtered from some insoluble material, concentrated to 10 ml, and refrigerated. Crystals of 1c (0.75 g) separated. The product was chromatographically pure and two recrystallizations from 50% aqueous ethanol gave pure 1c, clusters of needles, mp 106-108°.

Anal. Caled for $C_{12}H_{17}N_5O_5 \cdot H_2O$; C, 43.77; H, 5.82; N, 21.27, Found: C, 43.93; H, 6.21; N, 21.22.

Acknowledgment.—The anthors are indebted to Mr. **D.** Nobbs for his expert preparation of chemical interinediates.

Unnatural Amino Acids. II. Congeners of DL-3-Carboxy-4-methoxyphenylalanine

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Several studies on the use of amino acid residues to transport biologically active groups across cell membranes have been reported.²⁻⁶ In the main, however,

(1) Deceased.

the possibility that an amino acid derivative of a pharmacologically active moiety would penetrate to sites nut reached by the parent compound has not been explored. In the first paper of this series,⁶ the preparation and some of the biological properties were described of three new amino acids related both to tyrosine and aspirin. One of the compounds, pL-3-carboxy-4-methoxyphenylalarine (CMPA; 4. $\mathbb{R}^1 = 3$ -COOH: $R^{a} = 4$ -CH₃O; $R^{3} = H$) was found to be an orally effective analysic agent in animals and man but was poorly and creatically absorbed. It served, however, as a reference compound in the search for compounds which are better absorbed and at least as potent. The congeners of pL-3-carboxy-4-methoxyphenylalanine which were prepared and evaluated (Tables I and II) include isomers, homologs, and analogs. All were synthesized from acylamidomalonates as shown in Scheme I.





The pharmacological data in Table I show that small structural changes in DL3-carboxy-4-methoxyphenylalanine (CMPA) are accompanied by increased toxicity and attenuation or loss of analgetic activity. Replacement of the methoxy group by ethoxy or chloro and of the carboxy group by formyl or carboxamido results in considerable loss of analgetic activity. Substitution of the benzene nucleus of CMPA with a methyl group does not seem to have a markedly deleterious effect on analgetic activity but causes a great increase in toxicity. One of the isomers of CMPA, DL-3carboxy-2-methoxyphenylalanine, shows analgetic activity in the same range as CMPA but is more toxic.

⁽²⁾ E. F. Elliot, A. T. Faller, and C. R. Harington, J. Chem. Soc., 85 (1948).

 ⁽³⁾ J. H. Burckialter and V. C. Stevens, J. Am. Chem. Soc., 73, 56 (1951)
 (4) F. Bergel and J. A. Stock, J. Chem. Soc., 2409 (1954); P. Bergel,

^{V. C. E. Burnop, and J. A. Stock,} *ibid.*, 1223 (1955).
(5) I. F. Larionov, E. M. Shkodinskaya, V. I. Troosheikina, A. S. Khokh-

<sup>lov, O. S. Vasina, and M. A. Movikova, Lancet, 269, 169 (1959).
(6) F. Leonard, N. Wajngurt, W. Tschannen, and F. B. Block, J. Med.</sup> Chem., 8, 812 (1965).

Notes

TABLE I ANALGETIC ACTIVITY AND ACUTE TOXICITY



^a Average of reaction times determined at 15, 30, 45, and 60 min after administration of compounds. See ref 6 for a description of the test method.





^a E, ethanol; EA, ethyl acetate; I, isopropyl alcohol; R, reprecipitated from 0.1 N NaHCO₃ with 0.1 N HCl; W, water.

The carboxaldehyde (5, Table I) was prepared as a potential biological precursor of CMPA. Its considerably greater toxicity than that of CMPA but complete lack of analgetic activity suggests that while it is probably much better absorbed, it is very slowly, if at all, converted to CMPA in vivo. The amide analog of CMPA (7, Table 1) was prepared on the basis of the similarity of its relationship with CMPA to that of glutamine and glutamic acid. It was hoped that, like glutamine, it would be better absorbed than the related dicarboxylic acid7 and would provide higher blood and brain levels of an analgetically active species. The poor analgetic activity of this compound could be directly correlated with its very poor gastrointestinal absorption⁸ determined by the method of Hogben, et al.⁹ It is noteworthy that ring methyla-

(9) C. A. Hogben, D. J. Tocco, B. B. Brodie, and L. M. Schanker, J. Pharmacol. Exptl. Therap., 125, 279 (1955). tion in this series markedly enhances toxicity and analgetic activity. It would seem that ring methylation may have the effect of forming a lipophilic center which facilitates absorption by passive diffusion. The situation we envision for compounds of this type parallels that of soaps which also have lipophilic and ionic centers and are able to partition both between fat and aqueous phases.

Experimental Section¹⁰

o-Methoxybenzoic acid, o-ethoxybenzoic acid, 2-hydroxy-mtoluic acid, o-methoxybenzaldehyde, and p-chlorotoluene were obtained from Distillation Products Industries, Inc. 2-Hydroxy-p-toluic acid was purchased from K & K Laboratories, Inc. The toluic acids were esterified and methylated by standard reactions and gave products with physical constants in agreement with the published data: methyl 2-methoxy-mtoluate, bp 95° (1.2 mm), lit.¹¹ 114-116 (14 mm); methyl 2-

(11) R. Anschütz, H. Aschenberg, H. Keulært, F. Krone, K. Riepenkroger, and V. Zerbe, Ann., 442, 18 (1925).

⁽⁷⁾ P. Schwerin, S. P. Bessman, and H. Waelsch, J. Biol. Chem., 184, 37 (1950).

⁽⁸⁾ Private communication from Dr. P. Greengard.

⁽¹⁰⁾ Microanalyses were performed by Mr. J. Deonorine of these laboratories. Melting points (corrected) were taken on a Thomas-Hoover melting point apparatus.





^a B, benzenc; E, ethanol; EA, ethyl acetate; I, isopropyl alcohol; P, petroleum ether.

methoxy-p-toluate, bp 108-110° (1.4 mm), lit.11 137-139° (14 mm). p-Chlorotolnene was converted to 2-chloro-5-methylacetophenone, bp 108-109° (7 mm), n²⁵p 1.5420; lit.¹² bp 239-241)°, n²⁵D 1.5419. The latter gave, when oxidized with NaOBr¹³ instead of with KMnO4 as described in the literature, 14 an 87.5%yield of 6-chloro-m-tolnic acid (mp 157-158°, lit.4 mp 167° which was esterified by the method of Clinton and Laskowski¹⁵ to methyl 6-chloro-*m*-toluate, yield 84%, bp 79-80° (0.8 mm), n^{22} D 1.5338.

Anal. Calcd for C₉H₉ClO₂: C, 58.55; H, 4.91. Found: C, 58.20; H, 4.89.

o-Ethoxybenzoic acid, o-methoxy-m-tolnic acid and o-methoxybenzaldehyde were chloromethylated with a mixture of formaldehyde, concentrated HCl, and gaseous HCl utilizing the procedure described in the first article of this series.⁶ The products had physical constants and analytical data as follows: 5-chloromethyl-2-methoxy-m-tolnic acid, mp 186-187° (Anal. Calcd for C₀H₁₁ClO₃: C, 55.97; H, 5.15. Found: C, 55.80; H, 4.70); 5-chloromethyl-2-ethoxybenzoic acid, mp 51-52° (Anal. Calcd for C₁₀H₁₁ClO₃: C, 55.97; H, 5.15. Found: C, 55.90; H, 5.58); 5-chloromethyl-2-methoxybenzaldehyde, mp 75-76°, lit.¹⁶ mp 76°.

The chloro- and methoxytolnates were brominated with Nbromosuccinimide as described below for methyl 6-chloro-mtoluate. The bromination products decomposed when distilled and were therefore used without purification in the subsequent alkylation reactions.

Methyl 5-Bromomethyl-2-chlorobenzoate .--- N-Bromosuccininide (17.8 g, 0.1 mole) and 4 g of benzoyl peroxide were added to a solution of 18.5 g (0.1 mole) of methyl 6-chloro-m-toluate in 250 ml of CCl₄. The reaction mixture was stirred and refluxed for 1 hr and filtered. The filtrate was concentrated in vacuo, and the residual syrup was dissolved in 300 ml of benzene and chromatographed on a column of neutral alumina. The column was washed with benzene, the combined eluate and benzene washings were concentrated in vacuo, and the syrupy residue was freed of traces of solvent by prolonged warming in vacuo; yield 20 g (75%).

Alkylations of diethyl acetamidomalonate with 5-chloromethyl-2-methoxy-m-tolnic acid and 5-chloromethyl-2-ethoxybenzoic acid were carried out in absolute ethanol in the presence of 2 equiv of sodium ethoxide as described previously for alkylations with 5-chloromethyl-2-methoxybenzoic acid.⁶ The preparation of diethyl acetamido(3-carbomethoxy-4-chlorobenzyl)malonate illustrates the method utilized for alkylations with the bromomethyl esters and 5-chloromethyl-2-methoxybenzaldehyde. The carboxamidomalonates were prepared from carboxymalonates as illustrated by the preparation of diethyl formamido(3-carboxamido-4-methoxybenzyl)malonate. Physical data on the new malonates are reported in Table III.

Diethyl Acetamido(3-carbomethoxy-4-chlorobenzyl)malonate. -Diethyl acetamidomalonate (15.7 g, 0.072 mole) and 18.7 g (0.071 mole) of methyl 5-bromoniethyl-2-chlorobenzoate wcrc added to a cool solution of NaOEt prepared from 1.7 g (0.074 g-atom) of Na and 100 nd of ethanol. The resultant mixture was refluxed with stirring for 5 hr, filtered, and concentrated in vacuo. The solid residue was recrystallized from a benzenehexane mixture, cyclohexane, and finally from 2-propanol to yield 12.5 g (44%) of product, mp 112-114°

Diethyl Formamido(3-carboxamido-4-methoxybenzyl)malonate .--- Thionyl chloride (3.3 g, 10% excess) was added to a solution of 9.2 g (0.025 mole) of diethyl formamido(3-carboxy-4methoxybenzyl)malonate. The mixture was refluxed for 10 hr and concentrated in vacuo to remove excess SOCI₂. The residue was taken up in benzene and the solution was saturated with gaseous NH_3 . The product crystallized and was filtered off. The filter cake was washed with water and recrystallized from a mixture of ethyl acetate and hexane. The product melted at 130-131°, yield 7.0 g (80%).

The following examples illustrate the methods employed for the preparation of the acylamidomalonic acids, acylamida acids and amino acids (Table II).

 $Formamido ({\it 3-carboxy-4-methoxy-5-methylbenzyl}) malonic$ formamido(3-carboxy-4-methoxy-5-methylber-Acid. Diethyl zyl)malonate¹⁷ (12.0 g, 0.0315 mole) was refluxed for 4 hr with 50 ml of 10% NaDH. The reaction mixture was cooled and acidified to pH 3 with dilute HCl. The precipitate was filtered off and was recrystallized from a mixture of water and isopropyl alcohol: mp 175-177°, yield 4.65 g (45%). Anal. Calcd for C_{td}H_BNO₃: C, 51.70; H, 4.61; N, 4.30.

Found: C, 51.65; H, 5.16; N, 4.01.

Formamido(3-carboxamido-4-methoxybenzyl)malonic Acid. - Diethyl formamido(3-carboxamido-4-methoxybenzyl)malonate¹⁸ (14.0 g, 0.04 mole) was added to 32 ml of 2.5 N (0.08 mole) of Nat)H. The mixture was warmed in a boiling-water bath for 20 min to dissolve the malonate, cooled, and acidified to congo red with concentrated HCl. The crystalline precipitate melted at 184-185° after recrystallization from water: yield 15.1 g (61)%).

.tnal. Caled for CardbaN₂O₅: C, 50.25; H, 4.52; N, 9.03. Found: C, 49.90; H, 4.96; N, 8.86.

⁽¹²⁾ F. Mayer and W. Freund, Ber., 55, 1049 (1922).

⁽¹³⁾ The method employed for this oxidation was the same as that which was used for the preparation of β -naphthoic acid described by "Organic Synthesis," Coll. Vol. 11, John Wiley and Sons, Inc., New York, N. Y., 1943, p 428.

⁽¹⁴⁾ A. Claus, J. Prakt. Chem., [2] 46, 20 (1892).

¹¹⁵⁾ R. D. Clinton and S. C. Laskowski, J. Am. Chem. Soc., 70, 3135 i 1948)

⁽¹⁶⁾ W. Beek and I. N. Kaye, J. Ocg. Chem., 16, 1434 (1951).

⁽¹⁷⁾ Dietbyl acetanido(3-carboxy-4-methoxy-5-methylbenzyl)malomate was recovered unchanged after 70 for of reflux with 2 NHCL. When, lowever, it was refluxed with concentrated HCl for 70 br, not only ester hydrolysis but decarboxylation and other cleavage occurred with the formation of on-3-carboxy-5-methyltyrosine hydrochloride (2, Table II).

⁽¹⁸⁾ Several mnsuccessful attempts were made to selectively by dr dyzethe ester and acetamido groups of dietbyl acetamido(3-carboxamido-1methoxybenzyl)malonate with dilute HCl or NaOH. Mixtures of amidoand aminomonocarbicxylic and dicarboxylic acids were obtained as shown by elemental analyses and infraced spectra. Selective saponification of the ester groups of this compound was achieved with 2 equiv of NaOH as illustrated for its formamido homolog, but conditions could not be found for cleavage of die N-acetyl groop of the resolding N-acetyl-3-carboxamido-4-methoxyplanylanine withoo) sinolianeous hydrolysis of the aromatic carboxamide graaps.

DL-3-(3-Carboxy-4-ethoxyphenyl)alanine Hydrochloride.— Diethyl acetamido(3-carboxy-4-ethoxybenzyl)malonate (4 g, 0.01 mole) was refluxed with 25 ml of 1.2 N HCl for 19 hr. The reaction mixture was concentrated *in vacuo* and the crystalline residue was recrystallized from isopropyl alcohol; yield 2.02 g (69.0%).

DL-3-(3-Carboxy-2-methoxyphenyl)alanine Hydrochloride.— Diethyl acetamido(3-carbomethoxy-2-methoxybenzyl)malonate (7 g, 0.0177 mole) was hydrolyzed with 100 ml of 1.2 N HCl for 20 hr. Concentration of the reaction mixture *in vacuo* gave a crystalline residue which was recrystallized from a mixture of isopropyl and ethyl alcohol; yield 2.7 g (55%).

DL-3-(3-Carboxy-4-methoxy-*m*-tolyl)alanine Hydrate.—A mixture of 4.5 g (0.0138 mole) of formamido-(3-carboxy-4-methoxy-5-methylbenzyl)malonic acid and 50 ml of 50% ethanol was refluxed for 24 hr. The white crystalline compound which precipitated was filtered off, washed with water, and dissolved in 0.1 N NaHCO₃ solution. The bicarbonate solution was charcoaled, filtered, and acidified. The precipitate was filtered off, washed with water, and dried; yield 1.8 g (41%).

pl-3-(3-Carboxamido-N-formyl-4-methoxyphenyl)alanine. Formamido(3-carboxamido-4-methoxybenzyl)malonic acid (6 g, 0.0193 mole) was refluxed for 7 hr with 90 ml of 1:1 ethanolwater. The mixture was concentrated *in vacuo* and gave a syrupy residue which crystallized while standing under ether at ice-bath temperature. It melted at 118-119° after recrystallization from ethanol; yield 4 g (84%).

Anal. Caled for $\dot{C}_{12}H_{14}\dot{N_2}O_5$: \dot{C} , 54.13; H, 5.25; N, 10.52. Found: C, 54.43; H, 5.59; N, 9.92.

DL-3-(3-Carboxamido-4-methoxyphenyl)alanine Hydrochloride.—3-(3-Carboxamido-N-formyl-4-methoxyphenyl)alanine (4 g, 0.015 mole) was refluxed for 7 hr with 200 ml of 0.0995 N HCl. The reaction mixture was concentrated to dryness *in vacuo*. The oily residue crystallized under a mixture of etherethanol and was recrystallized from alcohol; yield 1.5 g (36.5%), mp 243-244°.

DL-3-(3-Carboxamido 4-methoxyphenyl)alanine.—Formamido-(3-carboxamido-4-methoxybenzyl)malonate (7 g, 0.022 mole) was refluxed for 26 hr in a mixture of 300 ml of 1:1 ethanolwater. The solvents were evaporated *in vacuo*, and the oily residue was taken up in the minimum amount of hot ethanol and allowed to crystallize in the refrigerator. The crystalline product was removed by filtration and recrystallized from ethanol; yield 3.5 g (66.6%), mp 269–270°.

pL-4-Carboxy-*m*-tyrosine Hydrate.—DL-3-(Carboxy-3-methoxyphenyl)alanine (3.0 g, 0.0125 mole) was refluxed with 30 ml of 48% HBr for 5 hr and allowed to cool overnight. The crystalline precipitate was filtered off and dissolved in water. The pH of the resulting solution was adjusted to 3.2 with 10% NaOH. The precipitate was filtered off, washed with water, and then dissolved in 0.1 N NaHCO₃. The bicarbonate solution was clarified (charcoal), acidified to pH 3.2, and filtered. The filter cake was washed well with water and dried; yield 1.55 g (51%).

Synthesis of Potential Antineoplastic Agents. XVII. N,N-Bis(2-fluoroethyl)anilines¹

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Prior to the report² of the clinical application of the drug ftorpan (I), essentially no work had been reported on the synthesis of 2-fluoroethylamines as potential



rationale for the synthesis of 2-fluoroethylamines has been presented.⁵ We report here the synthesis and some screening results on a number of substituted N,Nbis(2-fluoroethyl)anilines (II).

In an initial approach to the synthesis of II it was found that N,N-bis(2-chloroethyl)aniline (IV, R = H) and N,N-bis(2-chloroethyl)-*m*-toluidine (IV, R =3-CH₃) could be converted to the corresponding fluoroethylanilines (II) by refluxing with anhydrous potassium fluoride in methanol. However, attempts to extend the generality of this reaction to other chloroethylanilines (IV) failed.

As an alternative approach it was decided to attempt to replace the *p*-tolylsulfonyloxy grouping because of the ease with which they could be prepared and because several of the desired compounds had already been reported.⁷ These tosylates (V) were prepared by reaction of III with *p*-toluenesulfonyl chloride in the presence of pyridine; the new compounds so prepared are included in Table I.



In initial examples it was found that the tosylate could be readily replaced by fluorine by refluxing V with excess anhydrous KF in methanol. This displacement was accomplished without difficulty to give VI (\mathbf{R} = H, R = 3-CH₃, and R = 4-CH₃). These and other N.N-bis(2-fluoroethyl)anilines are included in Table II. However, when applied to V (R = 3-F and R = $3-NO_2$) and to VI this method led to the isolation of a N-(2-fluoroethyl)-N-(2-methoxyethyl)aniline (VII and VIII, respectively). These and other related compounds are included in Table III. Use of absolute ethanol in place of methanol with VI led to IX. Several other attempted displacements in methanol gave oils from which pure products could not be separated. Treatment of N,N-bis(2 fluoroethyl)aniline (IX) or compounds of the type VII with refluxing methanol or

^{(1) (}a) Part XVI: P. Schuyler, F. D. Popp, and A. C. Noble, J. Med. Chem., 9, 774 (1966). (b) Supported in part by research grants from the American Cancer Society and from the National Cancer Institute, U. S. Public Health Service.

^{(2) (}a) L. P. Larinov, M. D. Chadakova, and E. I. Arkhangel'skaia, Vopr. Onkol., 7, 112 (1961); Cancer Chemotherapy Abstr., 2, 872 (1961); (b) see also H. Dubicki, F. Zielinski, and F. W. Starks, J. Pharm. Sci., 53, 1422 (1964).

^{(3) (}a) A. P. Martinez, W. W. Lee, and L. Goodman, *Tetrahedron*, **20**, 2763 (1964); (b) A. P. Martinez, W. W. Lee, and L. Goodman, *J. Med. Chem.*, **8**, 741 (1965).

 ^{(4) (}a) Z. B. Papanastassiou and R. J. Bruni, J. Org. Chem., 29, 2870 (1964);
 (b) Z. B. Papanastassion, R. J. Bruni, F. P. Fernandes, and P. L. Levins, J. Med. Chem., 9, 357 (1966).

⁽⁵⁾ G. R. Pettit and R. L. Smith, Can. J. Chem., 42, 572 (1964).

⁽⁶⁾ Our work in this area began before these 1 eports^{3-5} appeared.

⁽⁷⁾ G. M. Timmis, British Patent 662,645 (1951).